**Faculty**

**Research Interests**

**Department of Biology**

**Villanova University**

**Revised: October 2022**

# Professor DR. ANIL BAMEZAI

Education

B.Sc. Jammu University, India 1979

M.Sc. Jammu University, India 1982

Ph.D. All India Institute of Medical Sciences, New Delhi, India 1987

Post-Doctoral Experience Harvard University 1987-1990

Dana-Farber Cancer Institute, Harvard University 1990-1995

Research

One research project in my laboratory focuses on the role of lipid rafts in spatiotemporal regulation of cell signaling in CD4+ T lymphocytes. Signaling cascades initiated by a number of cell surface proteins after binding their ligand are well characterized but a cohesive model that integrates signaling in space and time on the plasma membrane remains poorly understood. We are investigating the role of compositionally heterogeneous, nano-meter size, cholesterol-, saturated lipid-rich domains play an important role in determining the outcome of responses and cell fate in CD4+ T lymphocytes. Lipid rafts are heterogeneous in composition (George et.al., 2006; Bamezai & Kennedy, 2008) and coalesce during cellular interactions in the absence of a specific antigen (Kennedy et.al., 2011). This later observation goes against the current paradigm which suggests that antigen alone plays a central role in the organization (of signaling proteins) on the plasma membrane. We are currently examining the role antigen-independent raft coalescence on the plasma membrane of CD4+ T cell in the responses they generate after encountering a specific antigen.

Second project in my laboratory concerns Immunobiology of Ly-6 proteins. While studies with transgenic mice have allowed us to gain insights into the signaling role of Ly-6 proteins expressed on CD4+ T cells, the mechanism of such regulation remains unclear. How does this GPI-anchored protein lacking transmembrane and cytoplasmic domains communicate and signal to the cell interior during development of lymphocytes and while mounting a fitting response to an infection? Engaging a tail-less Ly-6 proteins on CD4+ T cell lines initiate signaling with wide array of functional consequences that range from the production of cytokines (IL-2), to growth inhibition and apoptosis. How does Ly-6A protein co-opts this membrane nano-structure and its contents (receptors, other signaling molecules) to communicate to the cell interior is not known. Membrane localization of Ly-6A protein to lipid rafts and its regulation of membrane proximal signaling event (Ca2+ flux experiments, Henderson et.al., 2002) provide a strong rationale for the focus on early membrane proximal events that involve lipid rafts. We are currently examining the lipid raft-dependent and independent signaling that occurs after engaging Ly-6 proteins.

Selected Publications

\*Lang, MA., Jenkins,SA., \*\*Balzano, P., \*Owoyele, A., \*Patel, A., and Bamezai, AK. 2017 Engaging Ly-6A/Sca-1 triggers lipid raft-dependent and -independent responses in CD4+ T-cell lines. Immunity, Inflammation and Disease 5 (4): 448-460. Online: 28 JUN 2017, DOI: 10.1002/iid3.182

\*Jones, M.,\*\*DeWolf, S., \*\*Vacharathit, V., \*\*Yim M., \*Spencer, S., and Bamezai, AK. 2016. Investigating B cell development, natural and primary antibody responses in Ly-6A/Sca-1 deficient mice PLOS ONE (In Press)

Comber, J.D and Bamezai, A. Gold Nanoparticles (AuNPs): A New Frontier in Vaccine Delivery. Journal of Nanomedicine Biotherapeutic Discov 2015, 5:4 (Invited Editorial)

\*Schieffer D, \*\*Naware S, \*Bakun W and Bamezai, AK. 2014. Lipid raft-based membrane order is important for antigen specific clonal expansion of CD4+ T lymphocytes. BMC Immunology, 15:58 (December 14, 2014). An "Editor's pick" article.

Bamezai AK and \*\*Divakar Lal 2014. Self-assembling nanoparticle: A strategy for designing universal flu vaccine. Journal of Nanomedicine and Biotherapeutic Discovery 4 (2): e129 (Invited Editorial)\*\*DeWolf, S, and Bamezai, A. 2013. Sex-specific effects of Stem cell antigen 1 (Sca-1)/Ly-6A in B lymphocyte development. (In Revision)

\*Comber J.D., and Bamezai A. 2012. In vitro derivation of interferon-¿ producing, IL-4 and IL-7 responsive memory-like CD4+ T cells. Vaccine, 30(12):2140-2145

\*Kennedy C, \*Nelson MD and AK. Bamezai. 2011. Analysis of Detergent-free Lipid Rafts isolated from CD4+ T cell line: Interaction with antigen presenting cells promotes coalescing of lipid rafts. BMC-Cell Communication and Signaling 9:31

Bamezai, A. 2008. "Membrane rafts and Signaling". Immunology, Endocrinology and Metabolic Agents in Medicinal Chemistry, (Invited Editorial) 8:325-326

\*Reed,J.,\*Branigan, P., and Bamezai, A. 2008. Interferon-gamma enhances clonal expansion and survival of CD4+ T cells. Journal of Interferon and Cytokine Research, 28: 611-618.

Bamezai, A., \*Kennedy, C. 2008. Cell-free antibody capture method for analysis of detergent-resistant membrane rafts. Methods in Molecular Biology, 477: 137-147.

\* Graduate student \*\* Undergraduate student

# Professor and Gerald M. Lemole Endowed Chair in Integrative Biology DR. AARON M. BAUER

Education

B. S. Michigan State University, East Lansing 1982

Ph.D. University of California, Berkeley 1986

Post-Doctoral Experience University of Calgary 1987-88

Research

My current research involves the analysis of evolutionary patterns in reptiles and amphibians. In particular I am interested in the phylogenetic systematics, evolutionary and functional morphology, and zoogeography of the geckos and other lizards of the southern hemisphere. Techniques used in this work include phylogenetic and phylogeographic techniques, DNA sequencing, x-ray CT-scanning, light and electron microscopy, whole body staining, radiography, and field research techniques.

Selected Publications

Čerňanský, A., Stanley, E.L., Daza, J.D., Bolet, A., Arias, S., Bauer, A.M., Vidal‑Garc, M., Bevitt, J.J., Peretti, A.M., Aung, N.N. and S.E. Evans. 2022. A new Early Cretaceous lizard in Myanmar amber with exceptionally preserved integument. Scientific Reports 12:1660. DOI: 10.1038/s41598-022-05735-5.

Bauer, A.M. and E.O. Lavilla. 2022. J.G. Schneider's *historiae amphibiorum*: herpetology at the dawn of the 19th century. Ithaca, New York: Society for the Study of Amphibians and Reptiles. 848 pp.

Stepanova, N.\* and A.M. Bauer. 2021. Phylogenetic history influences convergence for a specialized ecology: comparative skull morphology of African burrowing skinks (Squamata; Scincidae). BMC Ecology and Evolution (2021) 21:86. DOI: 10.1186/s12862-021-01821-w.

Bernstein, J.M.\*, Jackman, T.R., Sadlier, R.A., Wang Y. and A.M. Bauer. 2021. A novel dataset to identify the endemic herpetofauna of the New Caledonia biodiversity hotspot with DNA barcodes. Pacific Conservation Biology. DOI: 10.1071/PC20055.

Wood, Jr., P.L., Guo, X., Travers, S.L., Su, Y-C., Olsona, K.V., Bauer, A.M., Grismer, L.L., Siler, C.D., Moyle, R.G., Andersen, M.J. and R.M. Brown. 2020. Parachute geckos free fall into synonymy: Gekko phylogeny, and a new subgeneric classification, inferred from thousands of ultraconserved elements. Molecular Phylogenetics and Evolution 146: 106731. DOI: 10.1016/j.ympev.2020.106731.

Griffing, A.H., Gamble, T. and A.M. Bauer. 2020. Distinct patterns of pigment development underlie convergent hyperpigmentation between nocturnal and diurnal geckos (Squamata: Gekkota). BMC Evolutionary Biology 20:40. DOI: 10.1186/s12862-020-01604-9.

Weinell, J.L.\*, Branch, W.R., Colston, T.J., Jackman, T.R., Kuhn, A., Conradie, W. and A.M. Bauer. 2019. A species-level phylogeny of *Trachylepis* (Scincidae: Mabuyinae) provides insight into their reproductive mode evolution. Molecular Phylogenetics and Evolution 136:183–195.

Marques, M.P., Ceríaco, L.M.P., D.C. Blackburn, A.M. Bauer. 2018. Diversity and distribution of the amphibians and terrestrial reptiles of Angola: Atlas of historical and bibliographic records (1840–2017). Proceedings of the California Academy of Sciences 65, Suppl. II:1–501.

Roll, U., Feldman, A., Novosolov, M., Allison, A., Bauer, A.M …. and S. Meiri. 2017. The global distribution of tetrapods reveals a need for targeted reptile conservation. Nature Ecology & Evolution 1:1677–1682.

Paluh, D.J.\* and A.M. Bauer. 2017. Comparative skull anatomy of terrestrial and crevice-dwelling *Trachylepis*skinks (Squamata: Scincidae) with a survey of resources in scincid cranial osteology. PLoS ONE 12(9): e0184414.

Daza, J.D., Stanley, E.L., Wagner, P., Bauer, A.M. and D.A. Grimaldi. 2016. Mid-Cretaceous amber fossils illuminate the past diversity of tropical lizards. Science Advances 2, e1501080.

Bauer, A.M. 2013. Geckos: The Animal Answer Guide. Johns Hopkins University Press, Baltimore. 159 pp., 16 pp. pls.

Gamble, T., Greenbaum, E., Russell, A.P., Jackman, T.R., and Bauer, A.M. 2012. Repeated origin and loss of toepads in geckos. PLoS ONE 7(6): e39429. Doi:10.1371/journal.pone.0039429.

Russell, A.P. and Bauer, A.M. 2008. The appendicular locomotor apparatus of *Sphenodon* and normal-limbed squamates.  Pp. 1-466 *in* Gans, C., Gaunt, A., and Adler, K.K., eds. Biology of the Reptilia, vol. 21. Society for the Study of Amphibians and Reptiles, Ithaca, NY.

Lamb, T. and Bauer, A.M. 2006. Footprints in the sand: independent reduction of subdigital lamellae in the Namib-Kalahari burrowing geckos. Proc. Royal Society B 273:855-864.

Bauer, A. M. and Sadlier, R. A. 2000 The Herpetofauna of New Caledonia . Soc. Stud. Amphib. Ithaca. 310 pp.

Bauer, A. M. 1994. *Das Tierreich. Gekkonidae* (volume 1, Australia and the Pacific). Walter De Gruyter Publishers, Berlin. 306pp.

\*Graduate

# Assistant Professor DR. STEPHANIE M. CAMPOS

Education

B.S. University of Texas at Arlington 2012

Ph.D. Indiana University 2018

Post-Doctoral Experience Georgia State University, Center for Behavioral Neuroscience 2018-20

Research

My research aims to understand how chemicals control our social lives, from the chemicals that regulate behavior to the social contexts that motivate behavior. To examine the evolutionary roots of neural systems and behavior, I implement integrative and comparative approaches that span several disciplines, combining techniques from the behavioral neurosciences, evolutionary biology and chemical ecology. I explore and manipulate endocrine and exocrine chemical signaling systems to study how chemicals alter behavior in context-dependent ways, within individuals (neuroendocrine hormones and neurotransmitters) and between individuals (semiochemicals). *Sceloporus* lizards have specialized scent glands that secrete chemical signals important for competitive and sexual interactions. I am interested in the composition of these chemical signals, their impact on social behavior, and the physiological correlates that influence both. I conduct both field and lab studies with local populations, as well as in the American southwest and Mexico.

* Which physiological, environmental, and social factors influence the composition of and behavioral responses to chemical signals?
* How do hormones and semiochemicals impact territorial behavior?
* What role does vasotocin play in chemosensory-mediated social behavior of lizards?
* How does social status impact chemical communication in signalers and receivers?

Selected Publications

**Campos SM**, Erley A\*, Ashraf Z\* and Wilczynski W\*\*. 2022. Signaler’s vasotocin alters relationship between responder’s forebrain catecholamines and communication behavior in lizards (*Anolis carolinensis*). *Brain, Behavior and Evolution*. 97:184-196. DOI: 10.1159/000524217. Invited manuscript.

**Campos SM** and Belkasim SS\*. 2021. Chemosensory communication in lizards and a potential role for vasotocin in modulating social interactions. *Integrative and Comparative Biology*. 61(1): 205-220. DOI: 10.1093/icb/icab044. Invited manuscript.

**Campos SM**, Rojas V and Wilczynski W. 2020. Arginine vasotocin impacts chemosensory behavior during social interactions of *Anolis carolinensis* lizards. *Hormones and Behavior*. 124. DOI: 10.1016/j.yhbeh.2020.104772.

**Campos SM**, Pruett JA, Soini HA, Zúñiga-Vega JJ, Goldberg JK, Garcia CV, Hews DK, Novotny MV and Martins EP. 2020. Volatile fatty acid and aldehyde abundances evolve with behavior and habitat temperature in *Sceloporus* lizards. *Journal of Behavioral Ecology*. 31(4) 978-991. DOI: 10.1093/beheco/araa044.

**Campos SM**, Strauss C\* and Martins EP. 2017. In space and time: territorial animals are attracted to conspecific chemical cues. *Ethology*. 123(2), 136-144. DOI: 10.1111/eth.12582.

Romero-Diaz C, Xu C, **Campos SM**, Herrmann MA\*, Kusumi K, Hews, DK, and Martins EP. 2021. Brain transcriptomic responses of Yarrow’s spiny lizard, *Sceloporus jarrovii*, to conspecific visual or chemical signals. *Genes, Brain and Behavior*. 20(7):e12753. DOI: 10.1111/gbb.12753.

Romero-Diaz C, **Campos SM**, Herrmann MA\*, Lewis KN, Williams DR, Soini HA, Novotny MV, Hews DK and Martins EP. 2020. Structural identification, synthesis, and biological activity of two volatile cyclic dipeptides in a terrestrial vertebrate. *Scientific Reports*. 10:4303. DOI: 10.1038/s41598-020-61312-8.

Romero-Diaz C, **Campos SM**, Herrmann MA\*, Soini HA, Novotny MV, Hews, DK, and Martins EP. 2021. Composition and compound proportions affect the response to complex chemical signals in a spiny lizard. *Behavioral Ecology and Sociobiology*. DOI: 10.1007/s00265-021-02987-5.

Romero-Diaz C, Pruett JA, **Campos SM**, Ossip-Drahos AG, Zúñiga-Vega JJ, García CV, Hews DK and Martins EP. 2021. Evolutionary loss of a signaling color trait is linked to increased response to chemical stimuli. *Proceedings of the Royal Society B: Biological Sciences*. DOI: 10.1098/rspb.2021.0256.

Hill RZ, Hoffman B, Morita T, **Campos SM**, Lumpkin EA, Brem RB and Bautista DM. 2018. The signaling lipid sphingosine 1-phosphate regulates mechanical pain. *Elife*. 7, e33285. DOI: 10.7554/eLife.33285.001.

Pruett JA, Zúñiga-Vega JJ, **Campos SM**, Soini HA, Novotny MV, García CV, Martins EP and Hews DK. 2016. Evolutionary interactions between visual and chemical signals: Chemosignals compensate for the loss of a visual signal in male *Sceloporus* lizards. *Journal of Chemical Ecology*. 42(11), 1164-1174. DOI: 10.1007/s10886-016-0778-8.

\*Undergraduate student \*\*Posthumous coauthor

# Professor DR. SAMANTHA CHAPMAN

Education

B.S. The Pennsylvania State University 1998

M.S. Northern Arizona University 2002

Ph.D. Northern Arizona University 2005

Post-Doctoral Experience The Smithsonian Environmental Research Center 2005-07

Research

My team and I collaborate to understand how climate change, nutrient pollution, and rising sea levels alter coastal ecosystems in the WETFEET project. We use in the field experiments to see how resilient coastal ecosystems are to these global change factors. We work with land managers to plan climate adaptation strategies for the green strips that protect our world’s coasts. I am also interested in how anthropogenic alterations of biodiversity can impact ecosystems and the functions they provide. You can find more about my work here: [www.wetfeetproject.com](http://www.wetfeetproject.com) and <https://www1.villanova.edu/university/liberal-arts-sciences/scholarship/centers/cbest.html>

Selected Publications

Sturchio, M.A., J. Chieppa, S.K. Chapman, G. Canas\*, and M.J. Aspinwall. 2022. Temperature acclimation of leaf respiration differs between marsh and mangrove vegetation in a coastal wetland ecotone. Global Change Biology. <https://doi.org/10.1111/gcb.15938>

Chapman, S.K. I.C. Feller, G. Canas\*, M.A. Hayes, N. Dix, M. Hester, J. Morris, J.A. Langley. 2021*.* Mangrove growth responses to warming differ across a latitudinal gradient. Ecology.  <https://doi.org/10.1002/ecy.3320>

Adgie, T.A. S.K. Chapman. 2021. Salt marsh plant community Structure influences success of *Avicennia germinans* during poleward encroachment. Wetlands. <https://doi.org/10.1007/s13157-021-01463-0>

Geoghegan, E.\* J.A. Langley, and **S.K. Chapman**. 2021. A comparison of mangrove and marsh influences on soil respiration rates: a mesocosm study. Estuarine Coastal and Shelf Science. <https://doi.org/10.1016/j.ecss.2020.106877>

Chapman, S.K., M.A. Hayes, B. Kelly\*, J.A. Langley. 2019. What is the oxygen sensitivity of coastal wetland carbon mineralization? Biology Letters 15: 20180407.<http://dx.doi.org/10.1098/rsbl.2018.0407>

Coldren, G.A. Langley, J.A. Feller, I.C. and S.K. Chapman. 2019*.* Warming accelerates mangrove expansion and surface elevation gain in a subtropical wetland. Journal of Ecology. <https://doi.org/10.1111/1365-2745.13049>

Kittredge, H.A.\*\*, T. Cannone\*\*, J. Funk\*, S.K. Chapman*.*2018. Soil respiration and extracellular enzyme production respond differently across seasons to elevated temperatures. Plant and Soil. <https://doi.org/10.1007/s11104-018-3591-z>

Langley, J.A. S.K. Chapman and 17 other authors. 2018. Ambient changes exceed treatment effects on plant species abundance in global change experiments. Global Change Biology. <https://doi.org/10.1111/gcb.14442>

Barreto, C.R.\*, E. M. Morrissey, D.D. Wykoff, and **S.K. Chapman**. 2018. Co-occuring mangroves and salt marshes differ in microbial community composition. Wetlands. <https://doi.org/10.1007/s13157-018-0994-9>

Megonigal, P.A., **S.K. Chapman**, M. Kirwan. J.A. Langley, P. Dijkstra, S. Crooks. 2018.    Coastal Wetland Responses to Warmingin*A Blue Carbon Primer: The State of Coastalland Carbon Science, Practice and Policy*. by L. Windham-Myers, S. Crooks and T.G. Troxler. CRC Press

Doughty C.D.\* K. C. Cavanaugh, C.R. Hall, I.C. Feller, **S.K. Chapman**, 2017. Impacts of mangrove encroachment and mosquito impoundment management on coastal protection services. Hydrobiologia DOI 10.1007/s10750-017-3225-0

Chapman, S.K. K.A. Devine\*\*, C. Curran\*\*, R.O. Jones\*, and F.S. Gilliam. 2016*.* Impacts of soil nitrogen and carbon additions on forest understory communities with a nitrogen long-deposition history. Ecosystems 19: 142-154 DOI 10.1007/s10021-015-9922-5

Doughty, C.L. \* J. A. Langley, W.S. Walker, I.C. Feller, R. Schaub, and S. K. Chapman. 2015*.* Mangrove range expansion rapidly increases coastal wetland carbon storage. Estuaries and Coasts. DOI10.1007/x12237-015-9993-8

\* Graduate student \*\* undergraduate student

# Professor DR. ROBERT L. CURRY

Education

A.B. Dartmouth College, Hanover, NH 1979

M.S. University of Michigan, Ann Arbor 1981

Ph.D. University of Michigan, Ann Arbor 1987

Post-Doctoral Experience Archbold Biological Station, Lake Placid, FL 1987-90

Research

My research interests include behavior, ecology, and conservation, with an emphasis on field studies of marked birds to address questions concerning ecological influences on social organization, mating systems, and hybridization. We also work on the ecology of the uniquely herbivorous ant-acacia jumping spider, *Bagheera kiplingi*.

The largest component of my current research program focuses on the northward-moving hybrid zone between Black-capped and Carolina Chickadees in southeastern Pennsylvania. This work involves comparative of Carolina Chickadee populations in Chester and Berks Cos. and a predominantly Black-capped population in Schuylkill Co., as well as intensive field study at Hawk Mountain Sanctuary, which hosts a mixed population of both parental species and many hybrids. Recently, we also began examine the ways in which animal **personality** (consistent behavioral differences among individuals) and **cognitive ecology** might influence—and be affected by—the hybridization process.

A second area of ornithological research focuses on social ecology and conservation of island mockingbirds, allies, and selected other Neotropical birds.

Methods common to both areas of ornithological research include direct behavioral observation, color-banding, monitoring of demographic parameters such as survival and nest success, behavioral field experiments, and laboratory-based genetic analyses. We analyze spatial patterns using a Geographic Information System (GIS) and we use selected molecular markers (mtDNA and single nucleotide polymorphisms (SNPs)) to investigate species identity, parentage, and population structure. Most student projects have involved a combination of laboratory and field approaches. My students and I collaborate with ornithologists at Lehigh (cognitive ecology), Cornell and Colorado (evolutionary ecology), and the Academy of Natural Sciences (avian endoparasites).

Research on *B. kiplingi* has so far been involved periods of field research in southeastern Mexico, Costa Rica, and Panamá. This study links up with our biennial Field Ecology & Evolution class, which involves 2 weeks of field study in Central America (most recently, in Costa Rica); most students who have studied *B. kiplingi* have first completed the “FEE” course.

**Selected Publications (for access to PDF versions, visit** [**http://robertcurrylab.com/**](http://robertcurrylab.com/)**)**

DaCosta, J. M., M. J. Miller, J. L. Mortensen, J. M. Reed, R. L. Curry, and M. D. Sorenson. 2019. Phylogenomics clarifies biogeographic and evolutionary history, and conservation status of West Indian tremblers and thrashers (Aves: Mimidae). Molecular Phylogenetics and Evolution 136:196-205

Hager, S. B., …, R. L. Curry, et al. 2017. Continent-wide analysis of how urbanization affects bird-window collision mortality in North America. Biological Conservation 212, Part A:209–215

Kelemen, E. P.\*, K. E. Zusi#, and R. L. Curry. 2015. Song repertoire of the Carolina Chickadee in southeastern Pennsylvania. Wilson Journal of Ornithology, Wilson Journal of Ornithology 127:271-276

Taylor, S. A., R. L. Curry, T. A. White, V. Ferretti, and I. J. Lovette. 2014. Spatiotemporally consistent genomic signatures of reproductive isolation in a moving hybrid zone. Evolution 68:3066-3081

Taylor, S. A., W. Hochachka, T. A. White, V. Ferretti, R. L. Curry, and I. J. Lovette. 2014. Climate-mediated movement of an avian hybrid zone. Current Biology 24:671-676

Lovette, I. J., B. S. Arbogast, R. L. Curry, R. M. Zink, C. A. Botero, J. P. Sullivan, A. L. Talaba, R. B. Harris, D. R. Rubenstein, R. E. Ricklefs, and E. Bermingham. 2012. Phylogenetic relationships of the mockingbirds and thrashers (Aves: Mimidae). Molecular Phylogenetics and Evolution 63:219-229

Meehan, C. J.\*, E. J. Olson, M. W. Reudink, T. K. Kyser, and R. L. Curry. 2009. Herbivory in a spider through exploitation of an ant-plant mutualism. Current Biology, 19:R982-R893

\*Reudink, M. W., S. G. Mech, S. P. Mullen, and R. L. Curry. 2007. Structure and dynamics of the hybrid zone between Black-capped Chickadees (*Poecile atricapillus*) and Carolina Chickadees (*Poecile carolinensis*) in southeastern Pennsylvania. Auk 124:463-478

Curry, R. L., L. M. Rossano\*, and M. W. Reudink\*. 2007. Behavioral aspects of chickadee hybridization. Pages 95-110 in Ecology and behavior of chickadees and titmice: an integrated approach (K. Otter, Ed.). Oxford University Press, Oxford, England.

\*Reudink, M. W., S. G. Mech, and R. L. Curry. 2006. Extrapair paternity and mate choice in a chickadee hybrid zone. Behavioral Ecology 17:56-62 \* Graduate Student # Undergraduate student

# Associate Professor DR. ANGELA J. DI BENEDETTO

Education

B.S. SUNY at Binghamton, Binghamton, NY 1982

Ph.D. Cornell University, Ithaca, NY 1989

Postdoctoral experience University of Pennsylvania, Philadelphia PA 1990-1995

Research

My overall research interest is to understand how the fundamental processes of cell death, differentiation, and division are integrated and cross-regulated, so that normal development of the vertebrate embryo is achieved. I use the freshwater tropical zebrafish as a model system, and our lab centers around the investigation of the critical epigenetic transcriptional co-regulator Brd2 (bromodomain-containing 2), which is implicated in the control of all three processes, and is both a maternal and zygotic factor during development. We ask questions such as: 1) What is the expression pattern of Brd2 throughout development and in different tissues, and what clues to function do these patterns provide? 2) What are the upstream regulators and downstream transcriptional targets of Brd2, what cellular function do these targets accomplish, and how do targets change through developmental time? 3) What molecules interact with the Brd2 protein and under what conditions, and how do these interactions elucidate regulatory connections between cell death, differentiation and division? 4) What are the effects of overexpressing or knocking down expression of Brd2 on germline and embryonic development, and what does this imply about its epigenetic role in the context of the whole organism? Since Brd2 has a closely related paralog (Brd2b) in zebrafish, we also conduct comparative evolutionary studies to assess the level of functional divergence that has occurred after gene duplication. Experimental approaches entail molecular, cellular and genetic techniques including: Northern and Western blot analysis, in situ hybridization to RNA in tissues and zebrafish embryos, immunohistochemistry, confocal microscopy, chromatin-immunoprecipitation (ChIP), DNA sequencing and analysis, bioinformatics and data-mining, cloning and polymerase chain reaction (PCR), microinjection of zebrafish embryos, transgenic line construction, and phenotypic and behavioral analyses.

Selected Publications

Jalali, F., DiBenedetto, A.J., and Karlsson, J.O.M. 2018. Chilling causes perivitelline granule formation in activated zebrafish oocytes. **Cryobiology 81:** 210-213.

Murphy\*, T., Melville\*, H., Fradkin\*\*, E., Bistany\*,G., Branigan\*\*, G, Olson\*, K., Comstock\*\*, C., Hanby\*\*, H., Garbade\*\*, E., and DiBenedetto A.J. 2017. Knock-down of epigenetic transcriptional co-regulator Brd2a disrupts apoptosis and proper formation of the hindbrain and midbrain-hindbrain boundary (MHB) region in zebrafish. embryos in zebrafish. **Mechanisms of Development 146**: 10-30.

Desmond, M.E., J. E. Knepper, A. J. DiBenedetto, E. Malaugh\*, S.Callejo, R. Carretero, M.-I. Alonsoand A.Gato. 2014.  “Focal Adhesion Kinase as a Mechano-transducer during Rapid Brain Growth of the Chick Embryo” *Intl. J. Dev. Biol*.  **58**:35-43.doi: 10.1387/ijdb.130305md

DiBenedetto, AJ, Guinto\*, JB, Ebert\* TD, Bee\*\* KJ, Schmidt\*\* MM, and Jackman TR. 2008. Zebrafish *brd2a* and *brd2b* are paralogous members of the bromodomain-ET (BET) family of transcriptional coregulators that show structural and expression divergence. **BMC Develepmental Biology 8:** 39-58.

DiBenedetto, A. J., J. Klick Stoddard\*\*, and B. J. Glavan\*\*, 2001. Cloning and molecular characterization of a novel gene encoding a WD-repeat protein expressed in restricted areas of adult rat brain. **Gene 271:** 21-31.

Wang, S., A. J. DiBenedetto, and R. N. Pittman. 1997. Genes induced in programmed cell death of neuronal PC12 cells and developing sympathetic neurons *in vivo*. **Developmental Biology 188:** 322-336.

DiBenedetto, A. J., and R. N. Pittman. 1995. Death in the balance. **Perspectives on Developmental Neurobiology. 3:** 109-117.

Pittman, R. N., and A. J. DiBenedetto. 1995. Apoptosis of undifferentiated and terminally differentiated PC12 cells.@ In: *Cellular Aging and Cell Death.* (Holbrook, J. J., G. R. Martin, and R. A. Lockshin, eds.). John Wiley and Sons, Inc., New York, pp. 255-265.

Pittman, R. N., and A. J. DiBenedetto. 1995. PC12 cells overexpressing tissue plasminogen activator regenerate neurons to a greater extend and migrate faster than control cells in complex extracellular matrix. **Journal of Neurochemistry 64:** 566-575.

Pittman, R. N., S. Wang, A. J. DiBenedetto, and J. C. Mills. 1993. A system for characterizing cellular and molecular events in programmed neuronal cell death. **Journal of Neuroscience** 13: 3669-3680.

\*\*Undergraduate student \*Graduate student

**Professor, Assistant Chair DR. VIKRAM IYENGAR**

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**Education**

B.A. Stanford University 1993

Ph.D. Cornell University 2001

Post-Doctoral Experience Cornell University 2001-2002

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**Research**

My research involves studying the behavioral ecology of arthropods, with an emphasis on sexual selection in insects. Arthropods are the most abundant and diverse group in the animal kingdom, and they occupy nearly every ecological niche in marine, freshwater and terrestrial habitats. The extraordinary evolutionary success of arthropods can be partly attributed to the remarkable diversity of mating systems, and these fascinating creatures provide many wonderful opportunities to do both field and laboratory studies. Sexual selection is an important area of behavioral ecology that explains phenomena including exaggerated male traits, female mating preferences, precopulatory courtship signals, and postcopulatory sperm selection. I am primarily interested in the reproductive behavior of arthropods, particularly species that are sexually dimorphic – that is, where strong competition for mates has ultimately lead to divergence in the appearance of males and females. The main goal of my research program is to examine how the costs and benefits of precopulatory choice and postcopulatory selection shape the evolution of mating systems, and we study a variety of insects including moths, damselflies, and earwigs.

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**Selected Publications** (for access to PDF versions, visit my website: <http://viyengar.clasit.org/>)

Dodgen, R.E. & Iyengar, V.K. (2019) Weaponry, size and sex ratio affect spatial distribution within small and large groups of the maritime earwig (Anisolabis maritima). Ethology 125: 306-315.

Hack, N.L. & Iyengar, V.K. (2017) Big wigs and small wigs: time, sex, size and shelter affect cohabitation in the maritime earwig (Anisolabis maritima). PLoS One 12(10): e0185754.

Kendall-Bar, J.M. & Iyengar, V.K. (2017) Sexual selection by the seashore: the roles of body size and weaponry in mate choice and competition in the maritime earwig (Anisolabis maritima). Behavioral Ecology and Sociobiology 71(1): 8.

Egan, A.L., Hook, K.A., Reeve, H.K. & Iyengar, V.K. (2016) Polyandrous females provide sons with more competitive sperm: support for the sexy-sperm hypothesis in the rattlebox moth (*Utetheisa ornatrix*). *Evolution* 70(1): 72-81.

Conner, W.E. & Iyengar, V.K. (2016) Male Pheromones in Moths: Reproductive Isolation, Sexy Sons, and Good Genes. In Allison, J.D. and Cardé, R.T. (eds.), Pheromone Communication in Moths: Evolution, Behavior and Application, University of California Press, Berkeley, CA.

Walsh, J.T. & Iyengar, V.K. (2015) Win, lose, or draw: effects of residency, size, sex, and kinship on high-stakes larval contests in a moth. *Ethology* 121(8): 733-739.

Iyengar, V.K., Castle, T. & Mullen, S.P. (2014) Sympatric sexual signal divergence among *Calopteryx* damselflies is correlated with increased intra- and interspecific male-male aggression. *Behavioral Ecology and Sociobiology* 68(2): 275-282.

Kelly, C.A., Norbutus, A., Lagalante, A.F. & Iyengar, V.K. (2012) Male courtship pheromones indicate genetic quality in an arctiid moth (*Utetheisa ornatrix*). *Behavioral Ecology* 23(5): 1009-1014.

Iyengar, V.K. & Reeve, H.K. (2010) Z-linkage of female promiscuity genes in the moth *Utetheisa ornatrix*: support for the sexy sperm hypothesis? *Evolution* 64(5): 1267-1272.

Iyengar, V.K. (2009) Experience counts: females favor multiply-mated males over chemically-endowed virgins in a moth (*Utetheisa ornatrix*). *Behavioral Ecology and Sociobiology* 63(6): 847-855.

Iyengar, V.K. & Starks, B.D. (2008) Sexual selection in harems: male competition plays a larger role than female choice in an amphipod. *Behavioral Ecology* 19(3), 642-649.

Bezzerides, A.L., Iyengar, V.K. & Eisner, T. (2008) Female promiscuity does not lead to increased fertility or fecundity in an arctiid moth (*Utetheisa ornatrix*). *Journal of Insect Behavior* 21(4): 213-221.

Iyengar, V.K. & Eisner, T. (2004) Male indifference to female traits in an arctiid moth (*Utetheisa ornatrix*). *Ecological Entomology* 29(3), 281-284.

Iyengar, V.K., Reeve, H. K. & Eisner, T. (2002) Paternal inheritance of a female moth’s mating preference. *Nature* 419(6909), 830-832.

# Professor DR. TODD R. JACKMAN

Education

B.S. University of California, Davis 1987

Ph.D. University of California, Berkeley 1993

Postdoctoral Washington University, St. Louis 1994-99

Research

My research has focused on two areas in evolutionary genetics: 1. Reconstructing the history of populations representing different stages of speciation and 2. Using DNA sequence data in combination with other data to provide a robust historical framework for examining evolutionary processes. In my research, I have tried to apply innovative tests of phylogenetic patterns to molecular data in order to make reliable inferences of history. I have studied the evolutionary histories of both salamanders and lizards. My studies of western species of Plethodontid salamanders includes recent introductions of species, measures of gene flow between populations and the effects of combining morphological and molecular data for systematic studies. My postdoctoral research involved documenting and reconstructing parallel adaptive radiations in Caribbean anoline lizards using DNA sequences as well as morphological and ecological measures of habitat use.

Selected Publications

Agarwal I, Biswas S, Bauer A, Jackman T, et.al. (2017) Cryptic species, taxonomic inflation, or a bit of both? New species phenomenon in Sri Lanka as suggested by a phylogeny of dwarf geckos (Reptilia, Squamata, Gekkonidae, Cnemaspis). Systematics and Biodiversity.

Gamble T, Greenbaum E, Jackman T, et.al. (2017) Repeated evolution of digital adhesion in geckos: a reply to Harrington and Reeder. Journal of Evolutionary Biology.

Heinicke M, Jackman T, Bauer A. (2017) The measure of success: geographic isolation promotes diversification in Pachydactylus geckos. BMC Evolutionary Biology.

B R Karin, M Metallinou, J L Weinell, T R Jackman, A M Bauer (2016) Resolving the higher-order phylogenetic relationships of the circumtropical Mabuya group (Squamata: Scincidae): An out-of-Asia diversification. Molecular phylogenetics and Evolution . Elsevier.

Brennan I, Bauer A, Jackman T (2016) Mitochondrial introgression via ancient hybridization, and systematics of the Australian endemic pygopodid gecko genus Delma. Molecular Phylogenetics and Evolution.

Gamble T, Greenbaum E, Jackman T, Bauer A. (2015) Into the light : diurnality has evolved multiple times in geckos. Biological Journal of the Linnean Society.

Vile M, Kelman Wieder R, Živković T, Scott K, Vitt D, Hartsock J, Iosue C, Quinn J, Petix M, Fillingim H, Popma J, Dynarski K, Jackman T, Albright C, Wykoff D (2014). N2-fixation by methanotrophs sustains carbon and nitrogen accumulation in pristine peatlands. Biogeochemistry.

Bauer A, Masroor R, Titus-Mcquillan J, Heinicke M, Daza J, Jackman T. (2013). A preliminary phylogeny of the Palearctic naked-toed geckos (Reptilia: Squamata: Gekkonidae) with taxonomic implications. Zootaxa.

Wood P, Heinicke M, Jackman T, Bauer A. (2012). Phylogeny of Bent-toed Geckos (Cyrtodactylus) Reveals a West to East Pattern of Diversification. Molecular Phylogenetics and Evolution

W R Branch, A M Bauer, T R Jackman et al. (2011) A new species of the Pachydactylus weberi complex (Reptilia: Squamata: Gekkonidae) from the Namib-Rand Reserve, southern Namibia., 1-15. In Breviora.

E L Stanley, A M Bauer, T R Jackman et al. (2011) Between a rock and a hard polytomy: rapid radiation in the rupicolous girdled lizards (Squamata: Cordylidae)., 53-70. In Molecular Phylogenetics and Evolution 58 (1).

D M Portik, A M Bauer, T R Jackman (2011) Bridging the gap: western rock skinks (Trachylepis sulcata) have a short history in South Africa., 1744-1758. In Molecular ecology 20 (8).

S V Nielsen, A M Bauer, T R Jackman et al. (2011) New Zealand geckos (Diplodactylidae): Cryptic diversity in a post-Gondwanan lineage with trans-Tasman affinities., 1-22. In Molecular Phylogenetics and Evolution 59 (1).

Dibenedetto AJ, Guinto JB, Ebert TD, Bee KJ, Schmidt MM, Jackman T.R. 2008. Zebrafish brd2a and brd2b are paralogous members of the bromodomain-ET (BET) family of transcriptional coregulators that show structural and expression divergence. BMC Developmental Biology 10:8-39 PDF.

Smith, S.A., Sadlier, R.A., Bauer, A.M.,Austin, C.C., and Jackman, T.R. 2007. Molecular phylogeny of the scincid lizards of New Caledonia and adjacent areas: Evidence for a single origin of the endemic skinks of Tasmantis. Molecular Phylogenetics and Evolution 43: 1151 -1166.

# Associate Professor DR. ADAM LANGLEY

Education

B.S. North Carolina State University 1998

M.S. Northern Arizona University 2000

Ph.D. Northern Arizona University 2005

Post-Doc Experience The Smithsonian Environmental Research Center 2005-07

Research

I am interested in how ecosystems respond to, and may feedback to, global change. The most complex and uncertain questions regarding future ecosystems occur in the rhizosphere where plant roots interact intimately with the soil microbes to carry out the majority of terrestrial carbon and nutrient cycling. I use novel isotopic and gas exchange techniques to address these questions. I’m also working on a global change experiment in a brackish marsh on the Chesapeake Bay examining plant and microbial response to elevated CO2 and nitrogen pollution and sea level rise.

Selected Publications

Lu M, Herbert E, Langley JA, Kirwan M, Megonigal JP. 2019. Nitrogen status regulates morphological adaptation of marsh plants to elevated CO2. [*Nature Climate Change*](https://www.nature.com/articles/s41558-019-0582-x) 9, pages764–768.

Song J, Wan S, Piao S, Knapp AK, Classen AT, Vicca S, Ciais P, Hovenden MJ, Leuzinger S, Beier C, Langley JA, Kardol P. 2019. A meta-analysis of 1,119 manipulative experiments on terrestrial carbon-cycling responses to global change. [*Nature Ecology & Evolution*](https://www.nature.com/articles/s41559-019-0958-3). 19:1-2.

Hovenden MJ, Leuzinger S, Newton PCD, Fletcher A, Fatichi S, Luscher A, Reich PB, Andresen LC, Beier C, Blumenthal DM, Chiariello NR, Dukes JS, Kellner J, Hofmockel K, Niklaus PA, Song J, Wan J, Classen AT, and Langley JA. 2019. Globally consistent influences of seasonal precipitation limit grassland biomass response to CO2 [*Nature Plants*](https://www.nature.com/articles/s41477-018-0356-x) 5, pages 167–173.

Chapman SK, Hayes MA, Kelly B, & Langley JA. 2019. Exploring the oxygen sensitivity of wetland soil carbon mineralization. [*Biology Letters*](https://royalsocietypublishing.org/doi/full/10.1098/rsbl.2018.0407), *15*(1), 20180407.

GA Coldren, JA Langley, IC Feller, SK Chapman. (2019) Warming accelerates mangrove expansion and surface elevation gain in a subtropical wetland. [*Journal of Ecology*](https://besjournals.onlinelibrary.wiley.com/doi/pdf/10.1111/1365-2745.13049).

Langley JA, Chapman SK, La Pierre KJ, Avolio M, Bowman WD, Johnson DS, Isbell F, Wilcox KR, Foster BL, Hovenden MJ, Knapp AK, Koerner SE, Lortie CJ, Megonigal JP, Newton PCD, Reich PB, Smith MD, Suttle KB,  Tilman D. 2018. Ambient changes exceed treatment effects on plant species abundance in global change experiments [*Global Change Biology*](https://onlinelibrary.wiley.com/doi/10.1111/gcb.14442)*24(12):5668-79*. https://doi.org/10.1111/gcb.14442

Megonigal, J. P., Chapman, S., Langley, A., Crooks, S., Dijkstra, P., & Kirwan, M. 2018. Coastal Wetland Responses to Warming. In ***A Blue Carbon Primer*** (pp. 133-144). CRC Press.

\*\*Charbonneau B, Wootton L, Wnek J, Langley JA, Posner, M. 2017. Invasive sedge stabilized dunes more than native grass during Superstorm Sandy. *Journal of Applied Ecology*

\*\*Pastore MA, Megonigal JP, Langley JA. 2017 Greenhouse gas footprint of a brackish marsh in response to elevated CO2 and nitrogen. *Biogeochemistry* 133:73-87.

Fatichi, S, Leuzinger S, Paschalis A, Langley JA, Barraclough AD, Hovenden M. 2016. Partitioning direct and indirect effects discloses the response of water-limited ecosystems to elevated CO2. *PNAS* 113: 12757-12762.

\*\* Pastore MA, Megonigal JP, Langley JA. 2016. Elevated CO2 promotes long-term nitrogen accumulation only in combination with nitrogen addition. *Global Change Biology 22*: *391-403.*

\*\*Doughty CL, Langley JA, Walker WS, Shaub R, Feller IC, Chapman SK. 2015. Mangrove range expansion rapidly increases coastal wetland carbon storage. *Estuaries & Coasts* 1-12.

Langley JA, White HK, \*Palanivel RA, Shannon T, Chapman SK. 2015. Marsh plants mediate the influence of nitrogen fertilization on degradation of oil from the Deepwater Horizon spill. Ecosphere 6:art126.

\*Undergraduate author \*\*Graduate author

**Associate Professor DR. JOHN M. OLSON**

Education

A.B. Stanford University, Stanford, CA 1977

M.S. University of Michigan, Ann Arbor, MI 1983

Ph.D. University of Michigan, Ann Arbor, MI 1990

Post-Doctoral Experience Northeastern University, Boston, MA 1990-92

Research

Metabolic/muscle physiology and environmental physiology. Ongoing research includes investigation of oxidative stress and the functional and morphological maturation of effector tissues for thermogenesis and locomotion in birds and mammals, muscle function in vertebrates and invertebrates, and ecotoxicology. I use a variety of techniques in my research, and integrate research at the ecological, organismal and suborganismal levels of organization.

Selected Publications

Olson, J.M., C. Kearney\*, G. Rivera\*\*. 2013. Antioxidant enzymes: Acute and chronic responses to exercise-induced oxidative stress in gastrocnemius muscle of mice. Integr. Comp. Biol. (2013) 53 (suppl 1): e346.

Olson, J.M., K. Allport\*\*, P. Kealey\*\*, S McWilliams, and U. Bauschinger. 2011.Effect of Diet and Training on Ketone Body Metabolism in Starlings. Integr. Comp. Biol. (2011) 51 (suppl 1): e103.

Olson, J.M., A. Caragiulo\*, B. Czerwinski-Shields\*, and D. Soucier\*\*. 2010. Prolonged Cold Exposure in Young Quail: avUCP, Ultrastructure and Catabolic Capacities in Skeletal Muscle. Integr. Comp. Biol. (2010) 50 (suppl 1): e128.

Shea, R. E., J. M. Olson, and R. E. Ricklefs. 2007. Growth rate, protein accumulation, and catabolic enzyme activity of skeletal muscles of Galliform birds. Physiol. Biochem. Zoology80(3):306–316. 2007.

Dawson, W. R. and J. M. Olson. 2003. Thermogenic capacity and enzymatic activities in the winter-acclimatized dark-eyed junco (*Junco hyemalis*). J. Thermal Biol. 28:497-508.

Olson, J. M. 2001. Ontogeny of catabolic and morphological properties of skeletal muscle of the red-winged blackbird (*Agelaius phoeniceus*). J. Comp. Physiol. 171(7):527-542.

Krijgsveld, K. L., J. M. Olson, and R. E. Ricklefs. 2001. Catabolic capacity of the muscles of shorebird chicks: maturation of function in relation to body size. Physiol. Biochem. Zool. 74:250-260.

Olson, J. M., F. M. A. McNabb, M. S. Jablonski\*\*, and D. V. Ferris\*\*. 1999. Thyroid development in relation to the development of endothermy in the red-winged blackbird (*Agelaius phoeniceus*). Gen Comp. Endocrinol. 1999. 116:204-212.

Olson, J. M. and R. L. Marsh. 1998. Activation patterns and length changes in hindlimb muscles of the bullfrog (*Rana catesbeiana*) during jumping. J. Exp. Biol. 201:2763-2777.

McNabb, F. M. A., and J. M. Olson. 1996. Development of thermoregulation and its hormonal control in precocial and altricial birds. Poultry and Avian Biology Reviews 7:111-125.

Marsh, R. L. and J. M. Olson. 1994. Power output of scallop adductor muscle during contractions replicating the in vivo mechanical cycle. J. Exp. Biol. 193:139-156.

Olson, J. M. 1994. The ontogeny of shivering thermogenesis in the red-winged blackbird (*Agelaius phoeniceus*). J. Exp. Biol. 191:59-88.

Olson, J. M., and R. L. Marsh. 1993. Contractile properties of the striated adductor muscle in the bay scallop, *Argopecten irradians* at several temperatures. J. Exp. Biol. 176:175-193.

Marsh, R. L., J. M. Olson, and S. K. Guzik. 1992. Mechanical performance of scallop adductor muscle during swimming. Nature 357:411-413.

Olson, J. M. 1992. Growth, the development of endothermy, and the allocation of energy in red-winged blackbirds (*Agelaius phoeniceus*) during the nestling period. Physiol. Zool. 65:124-152.

Olson, J. M. 1991. Thermal relations of nestling red-winged blackbirds in southeastern Michigan. Auk 108:711-716.

Marsh, R. L., W. R. Dawson, J. J. Camilliere, and J. M. Olson. 1990. Regulation of glycolysis in the pectoralis muscles of seasonally acclimatized American goldfinches exposed to cold. Am. J. Physiol. 258 (Regulatory Integrative Comp. Physiol. 27):R711-R717.

Olson, J. M., and K. M. Crawford. 1989. Seasonal changes in buffering capacities and the activity of LDH in the heart and skeletal muscle of a vertebrate facultative anaerobe. J. Exp. Biol. 145:471-476.

\* Graduate student \*\* Undergraduate student

# Assistant Professor DR. DANA A. OPULENTE

Education

B.S. SUNY Stony Brook University 2006

Ph.D SUNY Stony Brook University 2015

Post-Doctoral Experience University of Wisconsin 2015-2020

Research

Budding yeasts have been isolated from soil, insects, plants, fruits, the ocean floor, and from every biome and continent. Furthermore, evidence suggests that ecological niches are partitioned by yeasts through temperature, host species, and sugar preferences. We will systematically explore the ecological, phenotypic, and genomic variation of species isolated across the Saccharomycotina subphylum, providing the opportunity to further explore the interplay between ecology and genetics of these species. Through a combination of sampling yeasts across multiple environments, high throughput phenotyping, and genomics, we will identify the traits important for different niches and the genetic differences responsible for these traits.

Selected Publications

Simone Mozzachiodi, F Bai, P Baldrian, P Buzzini, N Čadež, FC Riffo, S D, R Dimitrov, KJ Fisher, BR Gibson, D Gouliamova, D Greig, L Heistinger, CT Hittinger, V Koufopanou, M Jecmenica, CR Landry, T Mašínová, ES Naumova, **DA Opulente**, JJ Peña, U Petrovič, IJ Tsai, B Turchetti, P Villarreal, A Yurkov, G Liti, P Boynton. 2021. Yeasts from temperate forests. *Yeast*

William J Spurley\*, KJ Fisher\*, QK Langdon\*, KV Buh, M Jarzyna, MAB Haase, K Sylvester, RV Moriarty, D Rodriguez, A Sheddan, S Wright, L Sorlie, AB Hittinger, DA Opulente\*, CT Hittinger. 2021. Substrate, temperature, and geographical among nearly 2000 natural yeast isolates. *Yeast* <https://doi.org/10.1002/yea.3679> *\*First authors*

A. L. Labella, D.A. Opulente, J.L. Steenwyk, C.T. Hittinger, A. Rokas. 2019. Variation and selection on codon usage bias across an entire subphylum. PLoS Genetics 5(7): e1008304. https://doi.org/10.1371/journal.pgen.1008304

J.L. Steenwyk, D.A. Opulente, J. Kominek, X.X. Shen, X. Zhou, A.L. Labella, N.P. Bradley, B.F. Eichman, N. Čadež, D. Libkind, J. DeVirgilio, A.B. Hulfachor, C.P. Kurtzman, C.T. Hittinger, and A. Rokas. 2019. Extensive loss of cell cycle and DNA repair genes in an ancient lineage of bipolar budding yeasts. PLOS Biology 17(5): e3000255https://doi.org/10.1371/journal.pbio.3000255.

D.A. Opulente, Q.K. Langdon, K.V. Buh, M.A.B. Haase, K. Sylvester, R.V. Moriarty, M. Jarzyna, S.L. Considine, R.M. Schneider, C.T. Hittinger. 2019. Pathogenic budding yeasts isolated outside of clinical settings. FEMS Yeast Research. 19(3): foz032 <https://doi.org/10.1093/femsyr/foz032>.

J. Kominek, D.T. Doering, D.A. Opulente, X.X. Shen, X. Zhou, J. DeVirgilio, A.B. Hulfachor, C.P.

Kurtzman, C.T. 2019. Hittinger. Eukaryotic acquisition of a bacterial operon. Cell. 176(6):1356-1366.e10. doi: 10.1016/j.cell.2019.01.034.

X.X. Shen, D.A. Opulente, J. Kominek, X. Zhou, J.L. Steenwyk, K.V. Buh, M.A.B. Haase, J.H.

Wisecaver, M. Wang, D.T. Doering, J.T. Boudouris, R.M. Schneider, Q.K. Langdon, M. Ohkuma, R. Endoh, M. Takashima, R. Manabe, N. Čadež, D. Libkind, C.A. Rosa, J. DeVirgilio, A.B. Hulfachor, M. Groenewald, C.P. Kurtzman, C.T. Hittinger, A. Rokas. 2018. Tempo and mode of genome evolution in the budding yeast subphylum. Cell. 175(6): 1533-1545 e20 https://doi.org/10.1016/j.cell.2018.10.023

Opulente, D.A., E.J. Rollinson, C. Bernick-Roehr, A.B. Hulfachor, A. Rokas, C.P. Kurtzman, C.T. Hittinger. 2018. Factors driving metabolic diversity in the budding yeast subphylum. BMC Biology. 16(1): 26.

Opulente D.A., C.M. Morales, L.B. Carey, J.S. Rest. 2013. Coevolution trumps pleiotropy: Carbon assimilation traits are independent of metabolic network structure in budding yeast. PLOS One 8(1): e54403.

**Assistant Professor DR. MEGAN L. POVELONES**

Education

B.S. Loyola University in Maryland 2002

Ph.D Johns Hopkins University School of Medicine 2009

Post-Doctoral Experience University of Oxford 2008-10

Post-Doctoral Experience Imperial College London 2010-12

Research

Kinetoplastids are a group of single-celled eukaryotic parasites, some of which cause disease in humans and animals. All pathogenic kinetoplastids are transmitted by insect vectors, in which the parasites go through distinct developmental programs, including a stage adhered to insect tissue. We are using genome-wide approaches to identify and manipulate candidate proteins that may be involved in parasite adhesion to insects. Pathogenic parasites must also adapt their metabolism to survive in their different hosts. In some cases, these changes are accompanied by striking changes in mitochondrial shape, the mechanisms of which are completely unknown. I am interested exploring this process as model for the relationship between organelle structure and function.

Selected Publications

Filosa, John N, Corbett T Berry, Gordon Ruthel, Stephen M Beverley, Wesley C Warren, Chad Tomlinson, Peter J Myler, Elizabeth A Dudkin, Megan L Povelones, and Michael Povelones. “Dramatic Changes in Gene Expression in Different Forms of Crithidia fasciculata Reveal Potential Mechanisms for Insect-Specific Adhesion in Kinetoplastid Parasites.” PLoS Neglected Tropical Diseases 13, no. 7 (2019): e0007570.

Budzak, James, Louise E Kerry, Aris Aristodemou, Belinda S Hall, Kathrin Witmer, Manish Kushwaha, Carys Davies, Megan L Povelones, Jacquelyn R McDonald, and Aakash Sur. “Dynamic Colocalization of 2 Simultaneously Active VSG Expression Sites within a Single Expression-Site Body in Trypanosoma brucei.” Proceedings of the National Academy of Sciences 116, no. 33 (2019): 16561–70.

DiMaio, John, Gordon Ruthel, Joshua J Cannon, Madeline F Malfara, and Megan L Povelones. “The Single Mitochondrion of the Kinetoplastid Parasite Crithidia fasciculata Is a Dynamic Network.” PloS One 13, no. 12 (2018): e0202711.

Maree, Johannes Petrus, Megan Lindsay Povelones, David Johannes Clark, Gloria Rudenko, and Hugh-George Patterton. “Well-Positioned Nucleosomes Punctuate Polycistronic Pol II Transcription Units and Flank Silent VSG Gene Arrays in Trypanosoma brucei.” Epigenetics & Chromatin 10, no. 1 (2017): 1–21.

Povelones, Megan L. “Beyond Replication: Division and Segregation of Mitochondrial DNA in Kinetoplastids.” Molecular and Biochemical Parasitology 196, no. 1 (2014): 53–60.

Povelones, Megan L, Calvin Tiengwe, Eva Gluenz, Keith Gull, Paul T Englund, and Robert E Jensen. “Mitochondrial Shape and Function in Trypanosomes Requires the Outer Membrane Protein, TbLOK1.” Molecular Microbiology 87, no. 4 (2013): 713–29.

Povelones, Megan L, Eva Gluenz, Marcin Dembek, Keith Gull, and Gloria Rudenko. “Histone H1 Plays a Role in Heterochromatin Formation and VSG Expression Site Silencing in Trypanosoma brucei.” PLoS Pathogens 8, no. 11 (2012): e1003010.

Clayton, April M, Jennifer L Guler, Megan L Povelones, Eva Gluenz, Keith Gull, Terry K Smith, Robert E Jensen, and Paul T Englund. “Depletion of Mitochondrial Acyl Carrier Protein in Bloodstream-Form Trypanosoma brucei Causes a Kinetoplast Segregation Defect.” Eukaryotic Cell 10, no. 3 (2011): 286–92.

Gluenz, Eva, Megan L Povelones, Paul T Englund, and Keith Gull. “The Kinetoplast Duplication Cycle in Trypanosoma brucei Is Orchestrated by Cytoskeleton-Mediated Cell Morphogenesis.” Molecular and Cellular Biology 31, no. 5 (2011): 1012–21.

Lindsay, Megan E, Eva Gluenz, Keith Gull, and Paul T Englund. “A New Function of Trypanosoma brucei Mitochondrial Topoisomerase II Is to Maintain Kinetoplast DNA Network Topology.” Molecular Microbiology 70, no. 6 (2008): 1465–76.

# Professor DR. MICHAEL P. RUSSELL

Education

B.S. San Diego State University, San Diego, CA 1980

M.S. San Diego State University, San Diego, CA 1984

Ph.D. University of California, Berkeley 1990

Post-Doctoral Experience California Academy of Sciences, San Francisco, CA 1990-92

Research

My research lies at the intersection of marine invertebrate ecology, and fisheries and population biology. Currently I have an active program focusing on the population and larval ecology of commercially important estuarine and marine invertebrates. My work has both basic and applied significance. The kinds of questions I ask shed light on the ecology, life history, and population biology of the organisms I study. At the same time the data I gather are fundamental to the sustainable and intelligent management of these valuable natural resources. Focusing on applied problems often leads to an increase in the basic understanding of nature.

Selected Publications

Narvaez, C. A\*\*\*., A. J. Moura\*\*, D. F. Scutella\*\*, J. P. Cucchiara,\*\* A. Y. Stark, and M. P. Russell. 2022. Plasticity in fluctuating hydrodynamic conditions: tube foot regeneration in sea urchins. Journal of Experimental Biology **225**: jeb.242848

Stark, A. Y., C. A. Narvaez\*\*\*, and M. P. Russell. 2020. Adhesive plasticity among populations of purple sea urchin (Strongylocentrotus purpuratus). The Journal of Experimental Biology **223**: jeb228544

Narvaez, C. A.\*\*\*, A. M. Padovani\*\*, A. Y. Stark, and M. P. Russell. 2020. Plasticity in the purple sea urchin (Strongylocentrotus purpuratus): Tube feet regeneration and adhesive performance. Journal of Experimental Marine Biology and Ecology **528**: j.jembe.2020.151381

Russell, M. P., V. K. Gibbs\*\*\*, and E. Duwan\*\*. 2018. Bioerosion by pit-forming, temperate-reef sea urchins: History, rates and broader implications. Plos One **13**:e0191278.

Russell, M. P. and C. A. Narváez. 2016. Skeletal ossicles in echinoids are unreliable chronometers. Marine Biology. 163: 156-164

Haag, N.\*, M. P. Russell, and C. Hernandéz\*\*\*. 2016*.* Effects of spine damage and microhabitat on resource allocation of the purple sea urchin *Strongylocentrotus purpuratus* (Stimpson 1857). Journal of experimental marine biology and ecology.482: 106-117

Lützen, J., Å. Jespersen, and M. P. Russell. 2015. The Pacific clam *Nutricola tantilla* (Bivalvia: Veneridae) has separate sexes and makes use of brood protection and sperm storage. Journal of Molluscan Studies. 1–10. doi:10.1093/mollus/eyv015

Russell, M. P. 2013. Echinoderm responses to variation in salinity. Advances in Marine Biology. 66: 171 – 213.

Haag, N\*\*., M. P. Russell, C. Hernandéz\*\*\*, and N. R. Dollahon. 2013*.* Assessing fluorochrome-staining efficacy in the green sea urchin, *Strongylocentrotus droebachiensis* (Müller). Cahiers de Biologie Marine. 54: 625 – 631.

Clemente, S., J. C. Hernandéz, G. Montaño-Moctezuma, M. P. Russell, and T. A. Ebert. 2013. Predators of the sea urchin *Strongylocentrotus purpuratus* and the effects of spatial refuge on juvenile survival. Journal of Experimental Marine Biology and Ecology. *160:* 579-590

Elliot, L. F.\*\*, M. P. Russell & J.C. Hernández. 2013*.* Estimating Echinoid test volume from height and diameter measurements. Pp. 105-112, In: Echinoderms in a Changing World: Proceedings of 13th International Echinoderm Conference. C. Johnson editor.

Russell, M. P., T. A., Ebert, V. Garcia\*\*, and A. Bodnar. 2013*.* Field and laboratory growth estimates of the sea urchin *Lytechinus variegatus* in Bermuda. Pp. 133-140, In: Echinoderms in a Changing World: Proceedings of 13th International Echinoderm Conference. C. Johnson editor.

Ebert, T. A., J. C. Hernandéz, and M. P. Russell. 2012. Ocean conditions and bottom-up modifications of gonad development in the sea urchin Strongylocentrotus purpuratus over space and time. Marine Ecology Progress Series. *467:* 147-166.

Falese, L. E.\*, M. P. Russell, and N. R. Dollahon. 2011. Spermcasting of spermatozeugmata by the bivalves *Nutricola confusa* and *N. tantilla*. Invertebrate Biology. 130:334-343.

Ebert, T. A., J. C. Hernandéz, and M. P. Russell. 2010. Problems of the gonad index and what can be done: analysis of the purple sea urchin *Strongylocentrotus purpuratus*. Marine Biology. 158: 47-58

Hernandéz, J. C. and M. P. Russell. 2010. Does presence of substratum cavities affect allometric growth in the sea urchin *Strongylocentrotus purpuratus*? Journal of experimental biology. 213: 520-525

\*Graduate student \*\*Undergraduate student \*\*\*Postdoctoral Fellow

# Associate Professor DR. LOUISE A. RUSSO

Education

B.S. Villanova University, Villanova, PA 1983

Ph.D. Pennsylvania State University, Hershey 1987

Research

Cell biology and physiology. My research interests are primarily focused on regulatory mechanisms of signals that modulate uterine tissue growth and remodeling. We use an in vivo model system in the rodent to more specifically define patterns of regulation induced by hormones and other bioactive chemicals. Laboratory projects center on two main areas:

1) Estrogen induced uterine tissue inflammation activated via rapid receptor signaling pathways. The current focus is on G-protein coupled estrogen receptor (GPER) which is a ubiquitous membrane associated receptor that activates a variety of rapid signaling pathways including those involved in cell proliferation and survival. Utilizing vivo morpholino knockdown techniques in an immature rat ovariectomized rat model, we are currently working to assess how estrogen-induced inflammatory response is altered in the absence of GPER expression and to further characterize roles of this signaling protein in rapid uterine tissue response to estrogen.

2) Effects of xenoestrogenic environmental contaminants such as bisphenol A or DEHP or natural plant based estrogenic agents such as genistein found in soybeans on mammalian tissue biology and metabolic response. These bioactive compounds are implicated as causative agents in breast and prostate cancer, human infertility, and metabolic imbalance/obesity. Humans are exposed to these agents primarily through food and water sources such as with bisphenol A and genistein. In our current studies, we utilize either immature ovariectomized rat or adult ovariectomized mouse model organisms with chronic chemical treatments in the presence or absence of estrogen to assess complex biological effects and chemical response interactions on multiple morphological and biochemical endpoints in hormone responsive tissues including uterus, mammary gland, and liver.

With either project area experimental techniques include protein expression analysis via Western blot or ELISA, visualization of protein expression via immunohistochemistry with confocal microscopy, transmission electron microscopy to visualize ultrastructure changes in tissue morphology, and mRNA expression profiling via qPCR.

Selected Publications

Yudt, M. R., Russo, L.A., Berrodin, T. J. , Ellis,D. , Jelinsky, S.A. , Cohen, J. C. , Cooch, N., Unwalla, R. J., Fensome, A., Wrobel, J., Zhang, Z., Nagpal, S. and Winneker, R. C. Discovery of a Novel Mechanism of Steroid Receptor Antagonism:WAY-255348 Modulates Progesterone Receptor Cellular Localization, Phosphorylation and Promoter Interactions. Biochemical Pharmacology, 82: 1709-1719, 2011.

Russo, L. A., B. J. Peano, S. P. Trivedi, T. D. Cavalcanto, B. A. Olenchock, J. A. Caruso, A. R. Smolock, O. Vishnevsky, and R. M. Gardner. 2009. Regulated expression of matrix metalloproteinases, inflammatory mediators, and endometrial matrix remodeling by 17beta-estradiol in the immature rat uterus. Reproductive Biology and Endocrinology 7:124. (Open Access article)

Russo, L., Vishnevsky, O.\*\*, Caruso, J.\*\*, and Gardner, R. 2006. The role of inflammation in estrogen-induced extracellular matrix turnover and MMP regulation in the immature rat uterus. FASEB Journal, Vol. 20, No. 5 Part II.

Hafey, M\*., Russo, L.A., and Dollahon, N. Selective estrogen receptor modulators, tamoxifen and the raloxifene analogue LY117018, induce changes in uterine collagen matrix organization. FASEB Journal, 2003.

Peano, B.J.\* and Russo, L.A. 17β-estradiol differentially regulates MMP-3, 7, and 9 in the immature rat uterus. FASEB Journal, 2003.

Russo, L.A. and Olenchock, B.A.\*\* 2000. In Vivo regulation of matrilysin mRNA expression by 17β-estradiol in the immature rat uterus. Molecular Biology of the Cell, Vol. 11 Suppl.: 259a.

Russo, L. A., Calabro, S. P., Filler\*, T. A., Carey, D. J. and Gardner, R. M. 2001. In Vivo regulation of Syndecan-3 Expression in the Rat Uterus by 17β-Estradiol. Journal of Biological Chemistry, Vol. 276, pp. 686-692.

Morgan, H. E., B. H. L. Chua, and L. A. Russo. 1992. Protein synthesis and degradation. In: The Heart and Cardiovascular System, Second edition. H. A. Fozzard et al., eds. Raven Press Ltd., New York, pp. 1505-1524.

Russo, L. A., and H. E. Morgan. 1991. Effects of diabetes on cardiac protein metabolism. In: Diabetic Heart. N. Nagano and N. S. Dhalla, eds. Raven Press, New York, pp. 249-262.

\*Graduate student \*\* Undergraduate student

# Assistant Professor DR. TROY R. SHIRANGI

Education

B.Sc. State University of New York at Stonybrook 2000

Ph.D. Brown University, Providence, RI 2007

Post-Doctoral Experience University of WI, Madison; Janelia Research Campus/HHMI 2007-16

Research

The Shirangi laboratory is interested in how genes build neural circuits that guide innate animal behaviors. We use the courtship behaviors of *Drosophila* as a model system. Work in our laboratory integrates a variety of biological areas ranging from functional neuroscience, neuroanatomy and behavior to developmental genetics, and molecular and evolutionary biology. For more information, visit our laboratory website at shirangilab.com.

Selected Publications

Evolution of sexual size dimorphism in the wing musculature of Drosophila. Tracy CB\*, Nguyen J\*\*, Abraham R\*\*, Shirangi TR. PeerJ. 2020 Jan 17;8:e8360. doi: 10.7717/peerj.8360.

Court RC, Armstrong JD, Borner J, Card G, Costa M, Dickinson MH, Duch C, Korff W, Mann R, Merritt D, Murphey R, Namiki S, Seeds A, Shepherd D, Shirangi T, Simpson J, Truman J, Tuthill JC, Williams D. A systematic nomenclature for the Drosophila ventral nervous system. bioRxiv 2017, 122952.

Shirangi TR. Motor Control: Winging It with a Few Good Muscles. Curr Biol. 2017 Feb 6;27(3):R115-R116.

Shirangi TR, Wong AM, Truman JW, Stern DL. *Doublesex* Regulates the Connectivity of a Neural

Circuit Controlling *Drosophila* Male Courtship Song. *Dev Cell*. 2016 Jun 20;37(6):533-44.

Shirangi TR, Stern DL, Truman JW. Motor control of *Drosophila* courtship song.*, Cell Rep.* 2013

5(3):678-86.

# Assistant Professor DR. ALYSSA Y. STARK

Education

B.S. University of California, Davis 2006

Ph.D. University of Akron 2014

Post-Doctoral Experience University of Louisville 2014-17

Research

My research program uses an integrative approach to explore how environmental factors affect the morphology, performance, and behavior of biological organisms. I integrate laboratory and field-based methods rooted in biology, with analytical and theoretical methods from physics, chemistry, and material science. Currently I use ants, geckos, and sea urchins (with Dr. Michael Russell) to explore questions about the **functional morphology of adhesion**.

I have three major areas of interest:

**1.** Research on the adhesive systems of ants and geckos has principally focused the physical **mechanism** of adhesion. However, this has left gaps in our understanding of the chemical, material, and morphological characteristics of these systems. My collaborators and I fill these gaps using techniques such as mass spectrometry, NMR, and SEM.

**2.** Most biological adhesive systems are tested in controlled laboratory conditions. However, this approach often neglects whole organism **performance**, **behavior**, and relevant abiotic and biotic environmental factors. To address these complex interactions, I test static and dynamic adhesion of live ants and geckos in a variety of ecologically relevant conditions in the laboratory and in the field. I use these results to make predictions and test questions about the behavior, ecology, and evolution of these systems.

**3.** I am interested in the **application** of bio-inspired design and biomimicry to real-world problems. I apply the insights I glean from the mechanistic, performance, and behavioral attributes of natural adhesive systems to synthetic adhesive systems. I also find that focus on application provides important opportunities for collaboration within and outside of academia, interdisciplinary education, and service.

Selected Publications (for a complete list of publications, visit [www.alyssaystark.com](file:///C:/Users/Alyssa%20Y%20Stark/Desktop/www.alyssaystark.com)) \*\*Undergraduate student, \*Graduate student

Singla, S., D. Jain, C. M. Zoltowski\*\*, S. Voleti\*\*, **A. Y. Stark**, P. H. Niewiarowski & A. Dhinojwala. 2021. Direct evidence of acid-base interactions in gecko adhesion. *Science Advances.*In press.

Ringenwald, B. E.\*\*, E. C. Bogacki\*\*, C. A. Narvaez & **A. Y. Stark.** 2021. The effect of variable temperature, humidity, and substrate wettability on Gecko (*Gekko gecko*) locomotor performance and behavior. *Journal of Experimental Zoology Part A: Ecological and Integrative Physiology.*335: 454– 463. [Link](https://onlinelibrary.wiley.com/doi/10.1002/jez.2463)

Mitchell, C. T.\*, C. Balda Dayan, D-M\*. Drotlef, M. Sitti & **A. Y. Stark.** 2020. The effect of substrate wettability and modulus on gecko and gecko-inspired synthetic adhesion in variable temperature and humidity. *Scientific Reports*. 10(19748). [Link](https://www.nature.com/articles/s41598-020-76484-6#citeas)

**Stark, A. Y.**, C. A. Narvaez & M. P. Russell. 2020. Adhesive plasticity among populations of purple sea urchin (*Strongylocentrotus purpuratus*). *Journal of Experimental Biology*. 223: jeb228544. [Link](https://jeb.biologists.org/content/early/2020/06/24/jeb.228544.abstract)

**Stark, A. Y.** & S. P. Yanoviak. 2020. Adhesion and running speed of a tropical arboreal ant (*Cephalotes atratus*) on rough, narrow, and inclined substrates. *Integrative and Comparative Biology.* 60(4): 829–839. [Link](https://academic.oup.com/icb/article-abstract/doi/10.1093/icb/icaa078/5857132)

Narvaez, C. A., A. Padovani\*\*, **A. Y. Stark** & M. P. Russell. 2020. Plasticity in the purple sea urchin (*Strongylocentrotus purpuratus*): tube feet regeneration and adhesive performance. *Journal of Experimental Marine Biology and Ecology.*528: 151381.

Fernhaber, S. A. & **A. Y. Stark**. 2019. Biomimicry: new insights for entrepreneurship scholarship. *Journal of Business Venturing Insights.* 12(e00137).

Russell, A., **A. Y. Stark** & T. Higham. 2019. The integrative biology of gecko adhesion: historical review, current understanding and grand challenges. *Integrative and Comparative Biology.*59(1): 101-116. [Link](https://academic.oup.com/icb/article/59/1/101/5486592)

**Stark, A. Y**., H. R. Davis\* & W. K. Harrison\*\*. 2019. Shear adhesive performance of leaf-cutting ant workers (*Atta cephalotes*). *Biotropica.* 51(4):572-580.

**Stark, A. Y.**& C. T. Mitchell\*. 2019. Stick or slip: adhesive performance of geckos and gecko-inspired synthetics in wet environments. *Integrative and Comparative Biology.* 59(1): 214-226.

**Stark, A. Y.** & S. P. Yanoviak. 2018. Adhesion and running speed of a tropical arboreal ant (*Cephalotes atratus*) on wet substrates. *Royal Society Open Science*. 5(11):181540.

**Stark, A. Y.**, K. Arstingstall\*\* & S. P. Yanoviak. 2018. Adhesive performance of tropical arboreal ants varies with substrate temperature. *Journal of Experimental Biology.* 221(1): jeb171843.

**Stark, A. Y.**, B. J. Adams\*, J. Fredley\*\* & S. P. Yanoviak. 2017. Out on a limb: the thermal microenvironment of tropical arboreal ants. *Journal of Thermal Biology.* 69: 32-38.

**Stark, A. Y.**, M. Klittich\*, M. Sitti, P. H. Niewiarowski & A. Dhinojwala. 2016. The effect of temperature and humidity on adhesion of a gecko-inspired adhesive: implications for the natural system. *Scientific Reports*. 6(30936).

Walker, C. S., R. L. Ethington & **A. Y. Stark**. 2016. Who is your champion? A look at the structure and function of animals to help solve a problem. *Science and Children.* 53(9): 39-45.

Jain, D.\*, **A. Y. Stark**, P. H. Niewiarowski, T. Miyoshi & A. Dhinojwala. 2015. NMR spectroscopy reveals the presence and association of lipids and keratin in adhesive gecko setae. *Scientific Reports.* 3(9594).

Badge, I.\*, **A. Y. Stark**, E. L. Paoloni\*\*, P. H. Niewiarowski & A. Dhinojwala. 2014. The role of surface chemistry on adhesion and wetting of gecko toe pads. *Scientific Reports.* 4(6643).

**Stark, A. Y.**, I. Badge\*, N. A. Wucinich\*\*, T. W. Sullivan\*\*, P. H. Niewiarowski & A. Dhinojwala. 2013. Surface wettability plays a significant role in gecko adhesion underwater. *Proceedings of the National Academy of Sciences USA*. 110(16): 6340-6345.

# Professor DR. R. KELMAN WIEDER

Education

B.A. Amherst College, Amherst, MA 1974

M.A. University of Missouri, Columbia 1978

Ph.D. West Virginia University, Morgantown 1982

Post-Doctoral Experience West Virginia University, Morgantown 1982-84

Research

Ongoing research activities focus on the biogeochemistry and ecosystem ecology of boreal peatland ecosystems. These ecosystems collectively contain 1/3 of the world’s soil carbon, and have accumulated this carbon as peat over the past several thousand years. Our research strives to understand past, present, and future carbon cycling in these systems, especially in the face of natural and anthropogenic disturbances. Current research focuses on the impacts of enhanced nitrogen and sulfur acid deposition on peatlands resulting from ongoing oil sands development in the Fort McMurray area of Alberta and on nitrogen cycling after fire in Alberta peatlands.

Selected Publications

Wieder, R.K., D.H. Vitt, M.A. Vile, J.A. Graham\*, J.A. Hartsock\*, H. Fillingim, M. House, J.C. Quinn, K.D. Scott, M. Petix\*, K.J. McMillen. 2019. Experimental nitrogen addition alters structure and function of a boreal bog: Critical load and thresholds revealed. *Ecological Monographs*, DOI 10.1002/ecm.1371.

Hartsock\*, J.A., R.K. Wieder, M.A. Vile. 2019. Nitrogen retention by *Sphagnum fuscum* in laboratory mesocosms: Responses to experimentally added NH4+-N and NO3--N. *Wetlands* 39: 79-85.

Stuart\*, J.E.M, R.K. Wieder, M.A. Vile. 2018. Net nitrogen mineralization in Alberta Bog peat is insensitive to experimentally increased nitrogen deposition and time since wildfire. *Biogeochemistry* 138: 155-170.

Wieder, R.K., M.A. Vile, K.D. Scott, C.M. Albright, K. McMillen, D.H. Vitt, M. Fenn. 2016. Differential effects of high atmospheric N and S deposition on bog plant/lichen tissue and porewater chemistry across the Athabasca Oil Sands Region. *Environmental Science and Technology* 50: 12630-12640.

Wieder, R.K., M.A. Vile, C.M. Albright, K.D. Scott, D.H. Vitt, J.C. Quinn, M. Burke-Scoll\*. 2016. Effects of altered atmospheric nutrient deposition from Alberta oil sands development on *Sphagnum fuscum* growth and C, N, and S accumulation in peat. *Biogeochemistry* 129: 1-19.

Graham\*, J.A, J.A. Hartsock\*, D.H. Vitt, R.K. Wieder, and J.J. Gibson. 2015. Linkages between spatio-temporal patterns of environmental factors and distribution of plant assemblages across a boreal peatland complex. *Boreas* 45: 207-219

Shotyk, W., R. Belland, J. Duke, H. Kempter, M. Krachler, T. Noernberg, R. Pelletier, M. Vile, K. Wieder, C. Zaccone, and S. Zhang. 2014. *Sphagnum* mosses from twenty-one ombrotrophic bogs in the Athabasca Bituminous Sands region fail to reveal significant atmospheric contamination of “heavy metals.” *Environmental Science and Technology* 48: 12603-12611.

Vile, M.A., R.K. Wieder, T. Živković\*, K.D. Scott, D.H. Vitt,J.A. Hartsock\*, C.L. Iosue, J.C. Quinn,M. Petix\*, H. Fillingim\*, J.M.A. Popma\*, K.A. Dynarski\*\*, T.R. Jackman, C.M. Albright and D.D. Wykoff. 2014. N2-fixation by methanotrophs sustains carbon and nitrogen accumulation in peatlands. *Biogeochemistry* 121: 317-328.

Yu, Z., D.H. Vitt and R.K. Wieder. 2014. Continental fens as effective carbon sinks during the Holocene in western Canada. *The Holocene* 24: 1090-1104.

Benavides, J.C., D.H. Vitt and R.K. Wieder. 2013. The influence of climate change on recent peat accumulation patterns of *Distichia muscoides* cushion bogs in the high elevation tropical Andes of Colombia. *Journal of Geophysical Research-Biogeosciences* 118: 1627-1635.

Wieder, R.K., M.A. Vile, K.D. Scott, D.H. Vitt, E. Brault\*\*, M. Harris\*\* and S.B. Mowbray\*. 2012. Disturbance and the peatland carbon sink in the Oil Sands Administrative Area. Pages 13-22 in D.H. Vitt and J. Bhatti (eds.), Restoration and Reclamation of Boreal Ecosystems, Cambridge University Press.

House\*, M., D.H. Vitt and R.K. Wieder 2012. Plant community recovery on “minimum disturbance” petroleum sites compared to burned sites in bogs of northern Alberta. Pages 202-217 in D.H. Vitt and J. Bhatti (ed.), Restoration and Reclamation of Boreal Ecosystems, Cambridge University Press.

\*Graduate student

\*\*Undergraduate student

**Associate Professor DR. JAMES W. WILSON**

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**Education**

B.S. Bates College 1992

Ph. D. Columbia University 1998

Postdoctoral Yale University 1998-2000

Tulane University 2000-2005

Arizona State University Biodesign Institute 2006-2008

**Research**

Our lab works on two main areas:

1. Highly-conserved yet unexplored genes in bacteria (as a resource for improving bacterial engineering and vaccine design)
2. Cloning and transfer of large gene systems in bacteria (to study their evolution and application in synthetic biology and cellular engineering across species)

Unexplored genes: The genomics revolution has allowed us to sequence and decode virtually all known bacterial genomes.  This has allowed us to identify a certain category of genes that are highly-conserved across bacterial species yet are unexplored and remain totally unstudied.  Nothing is known about the role of these genes in bacterial biology.  We believe these genes represent an untapped resource of gene function that would be beneficial to bacterial engineering and help us learn about conserved gene mechanisms in different bacteria.  We target these unexplored regions of the bacterial genome for study.  We have found a variety of conserved phenotypes associated with these genes in Gram negative bacteria (mostly starting off in *Salmonella* but branching out to other species as well).  These discoveries establish new tools for bacterial engineering, synthetic biology, and vaccine design.

Cloning and transfer of large gene systems: Using techniques termed VEX-Capture and FRT-Capture, we are able to conveniently clone large genomic segments in bacteria (40-100+ Kb) and easily transfer these segments to new bacterial recipients.  We have targeted large contiguous gene systems for analysis using this approach.  We cloned the entire SPI-1 and SPI-2 type three secretion systems from *Salmonella enterica* and have studied their expression and function in other Gram negative recipients.  We have also cloned the entire *pdu/cob/cbi* gene system from *Salmonella enterica* (which functions to form a protein-based organelle called a microcompartment or MCP), and we have shown that this system is functional across different bacteria.  These studies open new doors to functional applications in bacteria and to new avenues of synthetic biology.

**Selected Publications**

Graf, L, Wu K, and **Wilson JW**. 2018. Transfer and analysis of Salmonella *pdu* genes in a range of Gram negative bacteria demonstrates exogenous microcompartment expression across a variety of species. Microbial Biotechnology. 2018 Jan;11(1):199-210.

Herman, A, Jacquelyn Serfecz, Alexandra Kinnally, Kathleen Crosby, Matthew Youngman, Dennis Wykoff, and **James W. Wilson**. 2016. The bacterial *iprA* gene is conserved across Enterobacteriaceae, involved in oxidative stress resistance, and influences gene expression in *Salmonella enterica* serovar Typhimurium. J. Bacteriol. 198(16): 2166-2179. Featured as Spotlight article.

Cangelosi C, Hannagan S, Santiago CP, and **Wilson JW**. 2015. Transfer of the cloned *Salmonella* SPI-1 type III secretion system and characterization of its expression mechanisms in Gram negative bacteria in comparison with cloned SPI-2. Microbiol Res. 2015 Nov;180:57-64.

Solomon L, Shah A, Hannagan S, and **Wilson JW**. 2014. Bacterial genus-specific tolerance for YdcI expression. Curr Microbiol. 2014 Nov;69(5):640-8.

Soni A, O'Sullivan L, Quick LN, Ott CM, Nickerson CA, and **Wilson JW**. 2014. Conservation of the Low-shear Modeled Microgravity Response in Enterobacteriaceae and Analysis of the *trp* Genes in this Response. Open Microbiol J. 2014 Jun 13;8:51-8.

Cangelosi, C., C. Shank, C.P. Santiago , and **J.W. Wilson**. 2013. Engineering large functional plasmids for biosafety. Plasmid 70 (2013) 385–392.

Jennings ME, Quick LN, Ubol N, Shrom S, Dollahon N, and **J.W. Wilson**. 2012. Characterization of Salmonella Type III Secretion Hyper-Activity Which Results in Biofilm-Like Cell Aggregation. PLoS ONE 7(3): e33080.

Santiago, C. P., L. N. Quick, and **J. W. Wilson**. 2011. Self-transmissible IncP R995 plasmids with alternative markers and utility for Flp/FRT cloning strategies. J. Microbiol. Biotechnol. 21(11):1123-1126.

Jennings M.E., Quick L.N., Soni A., Davis R.R., Crosby K., Ott C.M., Nickerson C.A., and **J.W. Wilson**. 2011. Characterization of the *Salmonella enterica* Serovar Typhimurium *ydcI* Gene, Which Encodes a Conserved DNA Binding Protein Required for Full Acid Stress Resistance. J. Bacteriol. 193(9):2208-17.

**Professor & Dennis M. Cook Endowed Gregor Mendel Chair in Genetics DR. DENNIS WYKOFF**

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**Education**

B.S. University of California, Davis 1993

Ph. D. Stanford University 1999

Postdoctoral University of California, San Francisco 1999-2005

Harvard University 2005-2006

**Research**

Our work studies how thiamine metabolism is regulated in *C. glabrata* at the level of thiamine biosynthetic promoters and transcriptional activators. By understanding how these promoters are regulated, rational strategies to alter the expression of thiamine starvation genes can be explored. Our work also examines whether thiamine nutrient status and the presence of *Cg*Pmu3 (and other thiamine regulated genes) influences the ability of *C. glabrata* to persist and multiply in phagocytic, macrophage-like cells. As clearance of *C. glabrata* is mediated by phagocytic cells. If the ability to acquire thiamine is defective in *C. glabrata*, then this fungal pathogen should be more easily cleared from the bloodstream by phagocytic cells. Long-term our work will provide a better understanding of the thiamine biosynthetic and acquisition pathway and determine if this pathway is a suitable target for the development of anti-fungal compounds.

**Selected Publications**

Bui LN@, Iosue CL, and Wykoff DD 2022. Tup1 paralog CgTUP11 is a stronger repressor of transcription than CgTUP1 in Candida glabrata***. mSphere*** 7(2) https://doi.org/10.1128/msphere.00765-21 (Editor’s pick)

Giuliano R, Roireau J, Rosano R, Lazzara N\*, Bajsa-Hischel J, Chen T, Schrader K, Duke S, & Wykoff D. 2020. Synthesis of Pyranopyrans Related to Diplopyrone and Evaluation as Antibacterials and Herbicides. ***Journal of Agricultural and Food Chemistry***. doi: 10.1021/acs.jafc.0c02564.

Iosue CL, Gulotta AP\*, Selhorst KB@, Mody AC@, Barbour KM@, Marcotte MJ@, Bui LN@, Leone SG\*, Lang EC@, Hughes GH@, & Wykoff DD. 2020. A Novel cis Element Achieves the Same Solution as an Ancestral cis Element During Thiamine Starvation in Candida glabrata. ***G3*** 10(1):321-331 doi: 10.1534/g3.119.400897

Rippel TM\*, Iosue CL, Succi PJ, Wykoff DD, & Chapman SK. 2020. Comparing the impacts of an invasive grass on nitrogen cycling and ammonia-oxidizing prokaryotes in high-nitrogen forests, open fields, and wetlands. ***Plant and Soil*** 449, pp. 65–77.

Patel K\*, Cangelosi C\*, Warrier V@, Wykoff D, & Wilson JW. 2020. The cloned SPI-1 type 3 secretion system can be functionally expressed outside Salmonella backgrounds. ***FEMS Microbiol Lett.*** 2020 367(8):fnaa065. doi: 10.1093/femsle/fnaa065.

Nahas JV@, Iosue CL, Shaik NF@, Selhorst K@, He BZ & Wykoff DD. 2018. Dynamic Changes in Yeast Phosphatase Families Allow for Specialization in Phosphate and Thiamine Starvation. ***G3*** pii: g3.200303.2018. doi: 10.1534/g3.118.200303.

Barreto CR, Morrissey EM, Wykoff DD & Chapman SK. 2018. Co-occurring mangroves and salt marshes differ in microbial community composition. ***Wetlands*** doi.org/10.1007/s13157-018-0994-9.

Burch J, Mashayekh S, Wykoff DD, & Grimes CL. 2018. Bacterial derived carbohydrates bind Cyr1 and trigger hyphal growth in *Candida albicans*. ***ACS Infectious Diseases*** 4(1):53-58. doi: 10.1021/acsinfecdis.7b00154.

Coldren GA, Barreto C#, Wykoff DD, Morrissey EM, Langley JA, Feller IC, & Chapman SK. 2016. Chronic warming stimulates growth of marsh grasses more than mangroves in a coastal wetland ecotone. Ecology 97(11):3167-3175. doi: 10.1002/ecy.1539.

Herman A#, Serfecz J#, Kinnally A@, Crosby K@, Youngman M, Wykoff D, & Wilson JW. 2016. The bacterial iprA gene is conserved across Enterobacteriaceae, involved in oxidative stress resistance, and influences gene expression in Salmonella enterica serovar Typhimurium. Journal of Bacteriology pii: JB.00144-16.

\* Graduate Student @=Undergraduate student

**Assistant Professor DR. MATTHEW J. YOUNGMAN**

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**Education**

B.S. Pepperdine University, Malibu, CA 1998

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Postdoctoral Massachusetts Institute of Technology, Cambridge, MA 2007-2012

**Research**

Work in my lab is focused on understanding the molecular basis of the changes in immunity that accompany aging,

including the age-dependent decline in immune function known as “immunosenescence”. We use a primarily

genetic approach to study host defense and the physiological response to infection during aging in the roundworm

*Caenorhabditis elegans*. Since many of the genetic determinants of lifespan were first discovered in *C. elegans*,

worms have become the preeminent model system in which to study the biology of aging. Moreover, worms are

protected from microbial infection by an ancient innate immune system consisting of antimicrobial peptides and

other immune effector proteins that are regulated by evolutionarily conserved immune signaling pathways. We

therefore anticipate that our discoveries will have significant implications for the underlying cause of deficient

immune function during aging in diverse species, including older humans. Ongoing work in the lab is directed

toward determining how the activity of immune signaling pathways is modulated during aging, identifying genes

with important roles in host defense later in life, and defining key age-dependent changes in cellular function that

impact resistance to infection.

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**Selected Publications**

Youngman, M.J., Rogers, Z.N., Kim, D.H. 2011. A decline in p38 MAPK signaling underlies immunosenescence in *Caenorhabditis elegans*. *PLoS Genetics 7(5):e1002082. doi:10.1371/journal.pgen.1002082.*

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Youngman, M.J. and Green, D.B. 1999. Microwave-assisted extraction of C60 and C70 from fullerene soot. *Talanta* 48:1203-1206.

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